Isolation of urease-positive *Ochrobactrum intermedium* in the stomach of a non-ulcer dyspeptic patient from north India

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*Ochrobactrum intermedium* is an opportunistic human pathogen found in immunocompromised individuals. We report the case of a north Indian patient with non-ulcer dyspepsia whose gastric biopsy revealed the presence of *O. intermedium*, along with *Helicobacter pylori*. Further description of *O. intermedium* was performed with 16S rRNA (1500 nucleotides) and RecA (1065 nucleotides) gene sequencing, and the identity and phylogenetic affiliation of the isolate was confirmed by 100% nucleotide similarity with *O. intermedium* LMG3301. Further investigation is required in order to evaluate the link between *H. pylori* and *O. intermedium* in the gastric niche.

**Key words:** Helicobacter pylori; Ochrobactrum; RecA recombinases; RNA, ribosomal, 16S

**Introduction**

The human stomach is colonized by diverse groups of bacteria, both pathogenic and non-pathogenic. Microorganisms in the stomach comprise a broad range of bacteria, including *Firmicutes*, *Proteobacteria*, *Actinobacteria* and *Fusobacteria*, streptococci and lactobacilli. Recently, it has been postulated that the stomach comprises 32 phylotypes from *Proteobacteria*, including 3 uncharacterized phylotypes [1]. *Helicobacter pylori*, a known causative agent of peptic ulcers and gastric cancer, colonizes the stomach of approximately 50% of the world’s population. However, little is known regarding the presence of bacteria other than *H. pylori* in the human stomach.

We describe for the first time the presence of *Ochrobactrum intermedium* in the antrum of a non-ulcer dyspepsia patient from India, who was diagnosed positive for *H. pylori*.

The genus *Ochrobactrum* comprises saprophytic, Gram-negative, oxidase-producing, lactose-non-fermenting, urease-positive, capsulating, aerobic bacilli belonging to alpha 2 subgroup of the class *Proteobacteria* (previously described as *Achromobacter* spp. or CDC group Vd), with 6 different species at present [2]. *Ochrobactrum* is a close genetic relative of the genus *Brucella*, as evidenced by protein profiling, Western blot, 16S rRNA and RecA sequencing-based studies, immunoelectrophoresis and amplified fragment length polymorphism [2-4]. *Ochrobactrum anthropi* has been the causative agent of various clinical and nosocomial outbreaks [3].

*O. intermedium* is an emerging pathogen in liver abscess post-liver transplantation and in bladder cancer patients [5]. Clinical manifestations and diseases caused by *O. intermedium* are poorly known and there are no suitable techniques for discriminating organisms up to species level in clinical cases. Conventional methods such as biochemical characteristics and antibiotic profiling do not provide accurate identification [5].

Recently, it has been shown that recombinase gene subunit A (*RecA*) is a reliable marker for the discrimination of *Ochrobactrum* taxon at inter- and intraspecies level [2]. We report the isolation of *O. intermedium* from the stomach of a male patient with non-ulcer dyspepsia.
Case Report

A patient with dyspeptic symptoms of bloating, gaseousness and postprandial epigastric pain of five to eight months’ duration underwent upper gastrointestinal endoscopy, as per the guidelines of the ethical committee at Motilal Nehru Medical College, India. The patient, designated as ‘M86’ in our study, was a 26-year-old male with non-ulcer dyspepsia. There was no history of medication except for occasional consumption of antacids. The patient denied taking non-steroidal anti-inflammatory drugs. He was vegetarian and did not consume alcohol. Clinical evaluation revealed mild epigastric tenderness. Ultrasound examination of the abdomen did not reveal any abnormality. The fasting and postprandial blood glucose, serum urea, bilirubin, amylase, transaminases and creatinine were within normal limits.

Upper gastrointestinal endoscopy did not reveal any abnormality. Antral biopsies were obtained and rapid urease test (HiMedia Laboratories, Mumbai, India) was positive. Hematoxylin and eosin-stained sections of antral biopsy showed focal areas of damage and mucin depletion of the epithelial lining. Gastric pits were mildly distorted and widely placed because of marked fibrosis, mainly in the upper part of the lamina propria. Gastric glands showed mild atrophy, mainly in the superficial areas. There was moderate infiltration by chronic inflammatory cells. Superficial mucosal capillaries were dilated and congested (Fig. 1). The organisms compatible with typical morphology of *H. pylori* were seen in the gastric pits. In addition, some Gram-negative short rods were visible. There was no evidence of neutrophil activity, intestinal metaplasia or lymphoid collection.

Polymerase chain reaction from the total biopsy DNA with *H. pylori*-specific primers [6] confirmed the presence of *H. pylori* by sequence analysis (Fig. 2). However, by standard culture approach using *Brucella* chocolate agar (Difco, Detroit, MI, USA) supplemented with *H. pylori*-selective supplement (Dent, Oxoid, Hampshire, UK), there were no colonies with typical *H. pylori* morphology, but colonies with pigmentation and morphology different from *H. pylori* were successfully isolated and subcultured. The colonies that appeared after 48 h incubation were urease-positive, oxidase-positive and non-hemolytic. Biochemical analysis of the isolate with API 20NE (bioMérieux, Marcy l’Etoile, France) and conventional tests identified the isolate to be of genus *Ochrobactrum* [2]. 16S
rDNA gene sequence of the isolate revealed 99% homology with *O. intermedium* LMG 3301 in GenBank. In phylogenetic analysis using Molecular Evolutionary Genetics Analysis (MEGA, Version 3.1; Institute of Molecular Evolutionary Genetics, University Park, PA, USA) [7], the isolate clustered with *O. intermedium* LMG3301 (Fig. 3). The 16S rRNA sequence of the isolate M86 deposited to GenBank under the accession number DQ100421.

Susceptibility to various antimicrobials, including beta-lactams, cephalosporins, macrolides, quinolones, sulfonamides and aminoglycosides was determined on Mueller-Hinton agar by using standard commercial antibiotic octadiscs (HiMedia Laboratories). Zones of inhibition were calculated as per the recommendations of the National Committee for Clinical Laboratory Standards, 2002. The isolate was sensitive to tetracycline (30 μg), trimethoprim-sulfamethoxazole (1.25/23.75 μg), tobramycin (10 μg), ciprofloxacin (5 μg) and ofloxacin (5 μg), and chloramphenicol (30 μg) and was resistant to 28 antibiotics, which included penicillin G (10 units), ampicillin (10 μg), cephalaxin (30 μg), erythromycin (15 μg), gentamicin (10 μg), metronidazole (5 μg), carbencillin (100 μg), trimethoprim (5 μg), norfloxacin (10 μg), imipenem (10 μg), kanamycin (30 μg), amikacin (30 μg) and amoxicillin-clavulanic acid (30 μg). The isolate was resistant to colistin (10 mg) and polymyxin B (300 units), indicating the authenticity and virulence of the species as *O. intermedium*.

**Discussion**

The human stomach harbors a diverse range of bacteria, ranging from non-pathogens to pathogens. However, the relationship between colonizing bacteria and clinical disease is frequently complex [8]. *Ochrobactrum* infection was first reported in the form of pancreatic abscess in 1980 [9]. *O. anthropi* and *O. intermedium* are well known opportunistic human pathogens causing bacteremia and sometimes serious diseases [3,10]. *O. anthropi* has been isolated from various clinical settings in immunocompetent and immunocompromised patients [11], including nosocomial sepsis in human immunodeficiency virus-positive patients [12], noma lesions, and septic shock and from infection complicating hemodialysis, vertebral osteomyelitis, urine, feces, wounds, throat and medical equipment. *O. intermedium* has been detected in post-liver transplant patients [13] and from clinical specimens [11]. Recently, *O. anthropi* was isolated from the stomach of squirrel monkeys (*Saimiri* sp.) which had developed gastritis [14].

We are not aware of any report in which *Ochrobactrum* spp. has been isolated from the stomach of human beings. A unique observation in the present
case report was evidence of severe fibrosis in the lamina propria of the gastric mucosa on histological examination of the gastric antral biopsy (Fig. 1). Whether or not this fibrosis was partially or totally due to O. intermedium is conjectural. In this context, it is interesting to note that O. anthropi has been shown to be associated with mild gastritis in squirrel monkeys. This case also had infection by H. pylori. Since the prevalence of H. pylori in the population is very high, whether or not it had a role in this patient’s symptoms and histological findings is not clear [15]. Bacteria other than H. pylori have been found in cases of chronic active gastritis [16]. There are reports of gastritis due to enterococci [17], and in some cases staphylococci have been shown to be associated with gastric disorders and isolated from antral biopsies [18] from patients with or without H. pylori colonization. Importantly, H. pylori and O. intermedium both produce urease, and thus the presence of Ochrobactrum may confound the urease test for H. pylori. The role of Ochrobactrum in gastric pathology remains uncertain and requires further investigation.

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References